# #615 Targeted Immune Cell Activation by Systemic Delivery of Toll-Like Receptor 9 Agonist **Antibody Conjugates Induce Potent Anti-Tumor Immunity**

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# Introduction

- Despite the success of checkpoint inhibitors and adoptive CAR-T therapies, only a fraction of cancer patients benefit. Novel therapies engaging both innate and adaptive immune response may produce more robust and durable anti-cancer immunity [1].
- Toll-like receptors (TLRs) play crucial roles in mounting potent innate immune responses against invading pathogens. Activation of TLR signaling leads to induction of inflammatory cytokines and priming of adaptive immunity [2].
- TLR9 agonists have shown clinical responses in melanoma patients by intratumoral injection [3].
- We developed a novel Toll-like Receptor Agonist Antibody Conjugate (TRAAC) platform to deliver a potent TLR9 agonist (T-CpG) for targeted immune activation via systemic administration.
- We evaluated various TRAAC approaches targeting either immune cell receptors or tumor specific antigens, demonstrating robust immune modulation and potent single agent anti-tumor activity in pre-clinical settings.

# Fig 1: TRAAC is a Toll-Like Receptor Agonist Antibody Conjugate platform intended for systemic delivery of T-CpG, a potent TLR9 agonist

Antibody: Versatile TRAAC targeting Specific immune cells, or Tumor microenvironment

Immune Activator: T-CpG Potent TLR9 agonist

- Linear, monomeric, and non-aggregated
- Sequence optimized for potency and stability
- Innovative design enables efficient site-specific conjugation



1. Kobold., Innate and adaptive immunity combined for cancer treatment. PNAS 2019, 116 (4) 1087-1088.

2. Dowling et al., Toll-like receptors: the swiss army knife of immunity and vaccine development Clin. Transl. Immunology. 2016 5(5), e85-10.

3. Hamid et al., Intratumoral Immunotherapy—Update 2019. The Oncol. 2019; 25:343-359.



Figure 2: NFκB reporter cells were stimulated with T-CpG or ODN2006 for 18hrs (A). Isolated human B cells were stimulated for 48hrs with B cell or myeloid TRAAC, surface marker expression was assayed by flow cytometry (B). Human, cynomolgus PBMCs, and mouse splenocytes were stimulated with B cell TRAAC, or T-CpG for 48hrs and expression of activating receptors were assayed by flow cytometry (C).

### Fig 3: TRAACs directed to human pDC, myeloid and B cells induce potent targeted cellular activation and cytokine production





Figure 3, Human PBMCs were stimulated for 48hrs in the presence of pDC, myeloid and B cell-directed TRAACs. Surface marker expression and cytokines was assayed by flow cytometry or ELISA.

## Fig 4: TRAACs elicit indirect activation of non-targeted immune cells



Figure 4. Human PBMCs were stimulated with myeloid TRAAC for 48hrs and assayed for cell markers of activation of non-targeted T cells and B cells by flow cytometry

# Immune cell targeting TRAAC







Figure 5. Mice bearing CT26 were intraperitoneally (i.p.) treated with PBS or myeloid TRAAC at 0.1mg/kg. 1mg/kg or 3mg/kg (A) or B cell TRAAC at 1mg/kg, 3mg/kg or 10mg/kg (B). Murine TRAACs are conjugated to mouse reactive T-CpG. Arrows indicate doses administrated.

# Fig 6: Durable anti-tumor memory response in tumor free syngeneic model following administration of immune cell targeted TRAAC



Figure 6. Tumor eradicated mice from MC38 bearing mice following 10mg/kg 2q3 systemic treatment with either myeloid TRAAC or B cell TRAAC were re-challenged 60-70 days post-initial tumor clearance. Naïve age-matched mice were used as control for tumor growth.

### Fig 7: TRAAC evokes rapid innate and adaptive immune signatures within tumor microenvironment



Figure 7. Mice bearing CT26 were dosed intraperitoneally (i.p.) once with B cell TRAAC or unconjugated antibody at 10mg/kg. Mouse reactive T-CpG was injected directly into the tumor at 50ug/mouse. Tumors were harvested 8hrs post-dose for nanostring analysis. Raw data was normalized and visualized using nsolver software by Nanostring







- models, as a single agent











activation

















# **Tumor Targeting TRAAC**

### Fig 8: HER2-directed TRAAC induces tumor antigen dependent myeloid



Figure 8. Bone marrow-derived mouse macrophages were co-cultured in the presence of HER2<sup>pos</sup> (A) or HER2<sup>neg</sup> tumors (B) and treated with HER2 TRAAC or mouse reactive T-CpG for 24hrs. Median MHCII was assessed by flow cytometry

### Fig 9: HER2-directed TRAAC demonstrates potent and durable anti-tumor response in syngenetic tumor model



Figure 9. Mice bearing HER2<sup>pos</sup> MC38 were intraperitoneally (i.p.) treated with HER2-directed TRAAC at 1mg/kg, 3mg/kg or 10mg/kg (A). Arrows indicate doses administrated. Tumor eradicated mice were rechallenged with HER2<sup>pos</sup>MC38 60 days post-initial tumor clearance, naïve mice were used as

# **Exploratory Studies in Cynomolgus Monkey**

 Immune targeting TRAACs were well tolerated following repeated intravenous injections in cynomolgus monkeys, as assessed by clinical observations, body weights, food consumption, hematology, serum chemistry and histopathology.

# Conclusions

 TRAAC is a novel immune activating platform that engages innate and adaptive immunity. • TRAAC delivers T-CpG, a potent differentiated TLR9 agonist, to targeted immune cells or tumor

microenvironment leading to immune activation.

TRAACs demonstrate robust and durable tumor regression in multiple syngeneic tumor

TRAACs exhibit favorable tolerability and safety profiles in cynomolgus monkeys.

Designed as a systemic therapy, TRAACs demonstrate potent targeted immune activation, robust anti-tumor activity and favorable tolerability in pre-clinical settings. TRAAC platform has broad therapeutic potentials against multiple solid tumor malignancies